

Mathematical Modelling of Pattern Formation in Yeast Biofilms

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About Me

- 2011–2015: UG/Honours, University of Adelaide
 - Free surface flow over topography
- 2016–2019: PhD, University of Adelaide
 - **Modelling yeast biofilm growth**



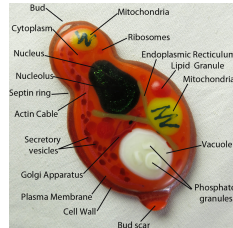
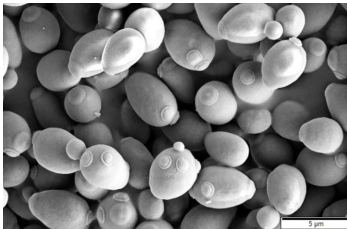
- Sep 2019–present: Postdoc, UQ
 - Modelling actomyosin networks in the cell cortex



- Twitter: [@xelamaths](#)

Yeast

- Single-cell fungi used in food and drink production (beer, wine, bread, vegemite)
- Bakers' yeast is a common model organism
 - Shares important characteristics with plant and animal cells
 - First eukaryotic genome to be completely sequenced
 - Helps develop antifungals and understand (cancer) cell division



Fungal Infections

- Pathogenic yeasts (e.g. *Candida albicans*) colonise medical devices and cause persistent infections
 - Resist antimicrobial therapy — expensive surgery often needed
 - Especially dangerous to immunocompromised people
 - Affects 1–2% of ICU patients, with up to 40% mortality rate¹
- Emerging pathogen *C. auris*: Japan 2009, 5 continents since
 - Highly resistant and difficult to diagnose

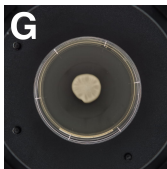


- We seek common mechanisms underlying yeast biofilm growth

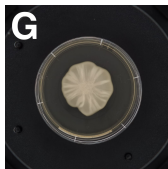
¹P. G. Pappas et al., *Nat. Rev. Dis. Primers* 4 (2018), 18026.

Yeast Biofilms

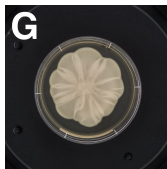
- To help them survive, pathogenic yeasts form **biofilms**: sticky communities of cells and fluid existing on surfaces
- Lab-grown biofilms of bakers' yeast form a floral pattern²



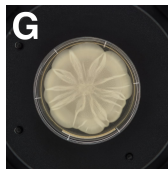
(a) Day 3



(b) Day 5



(c) Day 7



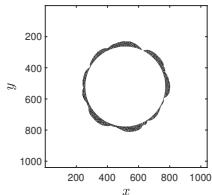
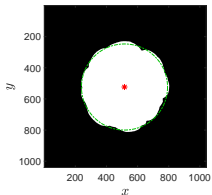
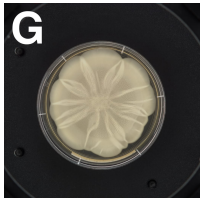
(d) Day 10

- Mechanisms of floral pattern formation only understood qualitatively
 - Nutrient-limited growth
 - Mechanical forces (e.g. extracellular fluid flow, adhesion, surface tension)

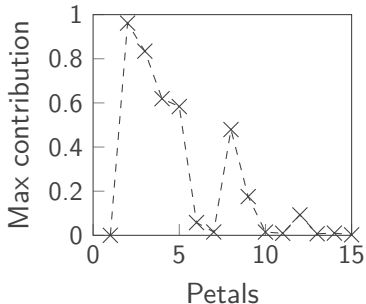
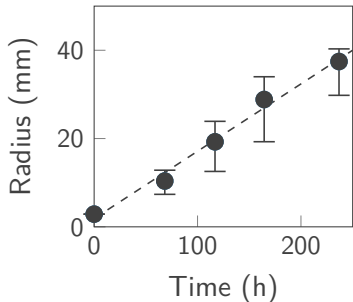
²T. B. Reynolds and G. R. Fink, *Science* 291 (2001), pp. 878–881.

Quantifying Biofilm Patterns

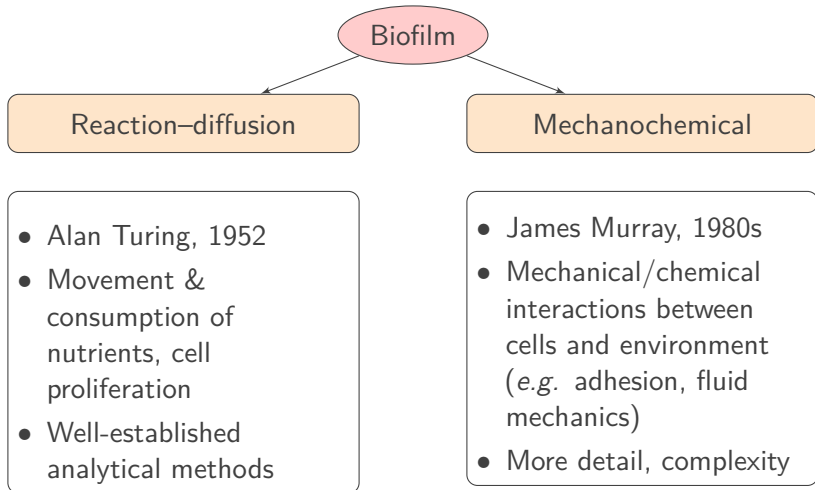
- We ran 13 experiments, and took 4 photographs of each



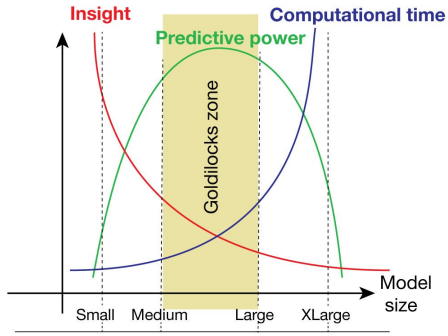
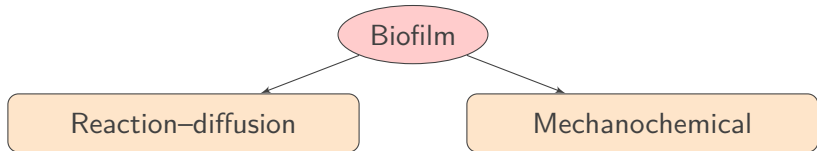
- Use spatial statistics to quantify biofilm size and shape



Modelling Pattern Formation



Modelling Pattern Formation



Nutrient-Limited Growth: Reaction–Diffusion Model

- Reaction–diffusion system with non-linear degenerate diffusion for cell spread
 - Enables cell density profiles with compact support
 - Models random motion of cells with non-unity aspect ratio³
 - $n(\mathbf{x}, t)$: numerical cell density
 - $g(\mathbf{x}, t)$: nutrient concentration
 - D : diffusion coefficient ratio, D_n/D_g
- Consider planar geometry accurate for $r \rightarrow \infty$

$$\frac{\partial n}{\partial t} = D \frac{\partial}{\partial x} \left(n \frac{\partial n}{\partial x} \right) + ng$$

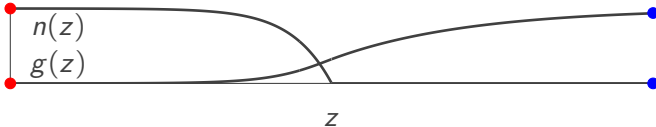
$$\frac{\partial g}{\partial t} = \frac{\partial^2 g}{\partial x^2} - ng$$

³M. J. Simpson, R. E. Baker, and S. W. McCue, *Phys. Rev. E* 83 (2011), 0121901.

Travelling Wave Analysis

- Travelling waves are a possible explanation for constant-speed expansion
- Introducing the travelling wave co-ordinates $z = x - ct$ and applying BCs yields a system of ODEs
- Defining $\zeta = \int_0^z n^{-1} ds$ removes singularity as $n \rightarrow 0$

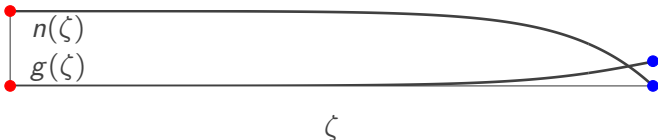
$$\begin{array}{l}
 n = 1 \\
 g = 0 \\
 w = 0
 \end{array}
 \quad
 \begin{array}{l}
 n \frac{dn}{dz} = \frac{1}{D} (c - cn - w - cg) \\
 \frac{dg}{dz} = w \\
 \frac{dw}{dz} = ng - cw
 \end{array}
 \quad
 \begin{array}{l}
 n = 0 \\
 g = 1 \\
 w = 0
 \end{array}$$



Travelling Wave Analysis

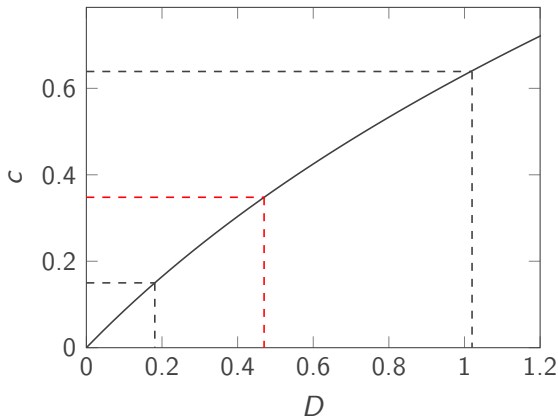
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$$\begin{array}{l}
 n = 1 \\
 g = 0 \\
 w = 0
 \end{array}
 \quad
 \begin{array}{l}
 \frac{dn}{d\zeta} = \frac{1}{D} (c - cn - w - cg) \\
 \frac{dg}{d\zeta} = nw \\
 \frac{dw}{d\zeta} = n^2 g - cnw
 \end{array}
 \quad
 \begin{array}{l}
 n = 0 \\
 g = g_0 \\
 w = 0
 \end{array}$$



Estimating the Diffusion Ratio

- There is a unique⁴ (minimum) wave speed c corresponding to each D
- We estimate D using experimental expansion speed
- Mean data: $D = 0.47$; Experimental range: $D \in [0.18, 1.02]$



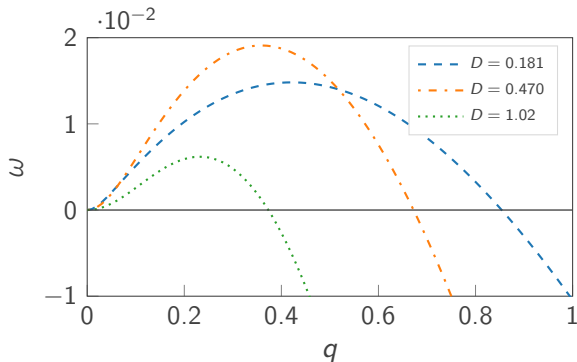
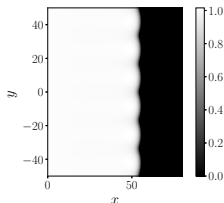
⁴J. Müller and W. van Saarloos, *Phys. Rev. E* 65 (2002), 061111.

2D Linear Stability Analysis

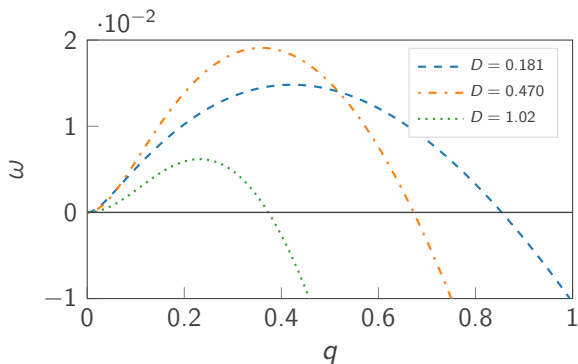
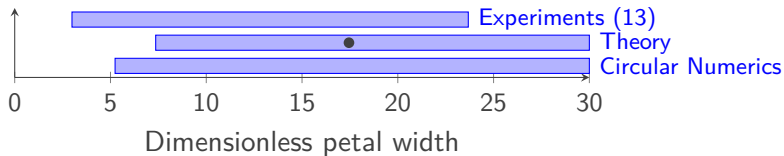
$$\xi = z + \varepsilon e^{iqy + \omega t}$$

$$n(\xi, y, t) = n_0(\xi) + \varepsilon n_1(\xi) e^{iqy + \omega t} + \mathcal{O}(\varepsilon^2)$$

$$g(\xi, y, t) = g_0(\xi) + \varepsilon g_1(\xi) e^{iqy + \omega t} + \mathcal{O}(\varepsilon^2)$$

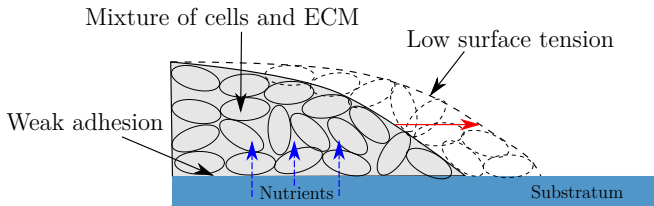


2D Linear Stability Analysis



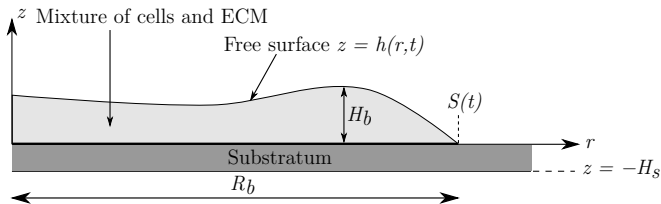
Sliding Motility

- We now consider mechanics in addition to nutrient limitation
- One hypothesis is that yeast biofilms expand by sliding motility⁵
 - Yeast adheres weakly to substratum — enables radial growth as cells proliferate
 - Biofilm takes up nutrients from the substratum
 - Nutrient consumption produces new cells and extracellular fluid
 - Cells and fluid spread passively as a unit



⁵T. B. Reynolds and G. R. Fink, *Science* 291 (2001), pp. 878–881.

Two-Phase Fluid Model



- Axisymmetric cylindrical geometry.
 - Biofilm occupies $0 \leq r \leq S(t)$ and $0 \leq z \leq h(r, t)$
- Biofilm is a mixture of two Newtonian viscous fluid phases:
 - Living cells $\phi_n(r, z, t)$ and ECM $\phi_m(r, z, t)$, with $\phi_n + \phi_m = 1$
 - Similar physical properties: $\rho_n = \rho_m$, $\mu_n = \mu_m$, etc.
 - Large interphase drag: $\mathbf{u}_n = \mathbf{u}_m$
- No tangential stress on biofilm–substratum interface
- Thin aspect ratio

$$\frac{H_s}{R_b} = \varepsilon \ll 1, \quad \frac{H_b}{R_b} = \mathcal{O}(\varepsilon)$$

Governing Equations

- Mass balance (fluid phases)

$$\frac{\partial \phi_n}{\partial t} + \nabla \cdot (\phi_n \mathbf{u}) = \psi_n \phi_n g_b - \psi_d \phi_n$$

$$\frac{\partial \phi_m}{\partial t} + \nabla \cdot (\phi_m \mathbf{u}) = \psi_m \phi_n g_b + \psi_d \phi_n$$

- Mass balance (nutrients in the **substratum** and **biofilm**)

$$\frac{\partial g_s}{\partial t} = D_s \nabla^2 g_s$$

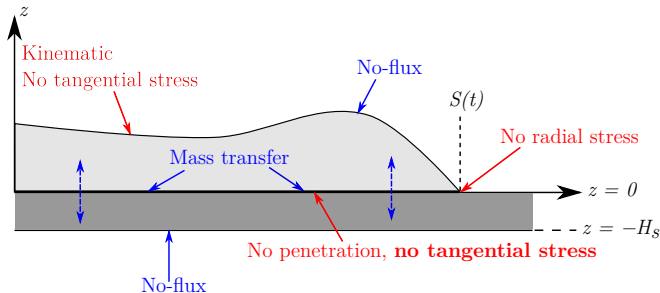
$$\frac{\partial g_b}{\partial t} + \nabla \cdot (g_b \phi_m \mathbf{u}) = D_b \nabla^2 g_b - \eta \phi_n g_b$$

- Momentum balance (fluid mixture)

$$\nabla \cdot \boldsymbol{\sigma} = \mathbf{0}$$

Boundary Conditions

- Boundary conditions for **nutrients** and **fluids** close the model



- Nutrient transfer conditions on $z = 0$:

$$D_s \frac{\partial g_s}{\partial z} = -Q (g_s - g_b), \quad D_b \frac{\partial g_b}{\partial z} = -Q (g_s - g_b)$$

- No tangential stress on the substratum models weak adhesion
- Free surface normal stress proportional to local curvature:

$$\hat{\mathbf{n}} \cdot (\phi_\alpha \boldsymbol{\sigma} \cdot \hat{\mathbf{n}}) = -\gamma \kappa \quad \text{on} \quad z = h$$

Extensional Flow Scaling

- Scaling based on relevant physics
 - Thin biofilm (aspect ratio $\varepsilon \ll 1$)
 - Low surface tension
 - Nutrient-limited growth
- Variables

$$(r, z) = (R_b \hat{r}, \varepsilon R_b \hat{z}), \quad (u_r, u_z) = (\psi_n G R_b \hat{u}_r, \varepsilon \psi_n G R_b \hat{u}_z),$$
$$t = \frac{\hat{t}}{\psi_n G}, \quad g_s = G \hat{g}_s, \quad g_b = G \hat{g}_b, \quad p = \psi_n G \mu \hat{p}$$

- Parameters (estimated based on experiments)

$$\Psi_m = \frac{\psi_m}{\psi_n} = 0.11, \quad \Psi_d = \frac{\psi_d G}{\psi_n} = 0, \quad \gamma^* = \frac{\varepsilon \gamma}{\psi_n G R_b \mu} = 0,$$
$$D = \frac{D_s}{\psi_n G R_b^2} = 4.34, \quad \text{Pe} = \frac{\psi_n G R_b^2}{D_b} = 0.95, \quad \Upsilon = \frac{\eta R_b^2}{D_b} = 3.15,$$
$$Q_s = \frac{Q R_b}{\varepsilon D_s} = 2.09, \quad Q_b = \frac{Q R_b}{\varepsilon D_b} = 8.65$$

Thin-Film Model

- Expand variables

$$h \sim h_0(r, t) + \varepsilon^2 h_1(r, t), \quad \phi_n \sim \phi_{n0}(r, z, t) + \varepsilon^2 \phi_{n1}(r, z, t), \quad \text{etc.}$$

- Dimensionless model (dropping hats)

$$\frac{1}{r} \frac{\partial}{\partial r} (ru_r) + \frac{\partial u_z}{\partial z} = (1 + \Psi_m) \phi_n g_b$$

$$\frac{\partial \phi_n}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} (ru_r \phi_n) + \frac{\partial}{\partial z} (u_z \phi_n) = \phi_n g_b - \Psi_d \phi_n$$

$$\frac{\partial g_s}{\partial t} = D \left[\frac{1}{r} \frac{\partial}{\partial r} \left(r \frac{\partial g_s}{\partial r} \right) + \frac{1}{\varepsilon^2} \frac{\partial^2 g_s}{\partial z^2} \right]$$

$$\text{Pe} \left(\frac{\partial g_b}{\partial t} + \nabla \cdot [(1 - \phi_n) g_b \mathbf{u}] \right) = \frac{1}{r} \frac{\partial}{\partial r} \left(r \frac{\partial g_b}{\partial r} \right) + \frac{1}{\varepsilon^2} \frac{\partial^2 g_b}{\partial z^2} - \Upsilon \phi_n g_b$$

$$-\frac{\partial p}{\partial r} + \frac{2}{r} \frac{\partial}{\partial r} \left(r \frac{\partial u_r}{\partial r} \right) - \frac{2}{3} \frac{\partial}{\partial r} \left[\frac{1}{r} \frac{\partial}{\partial r} (ru_r) + \frac{\partial u_z}{\partial z} \right] + \frac{\partial}{\partial z} \left(\frac{\partial u_z}{\partial r} + \frac{1}{\varepsilon^2} \frac{\partial u_r}{\partial z} \right) - \frac{2}{r^2} u_r = 0$$

$$-\frac{\partial p}{\partial z} + 2 \frac{\partial^2 u_z}{\partial z^2} - \frac{2}{3} \frac{\partial}{\partial z} \left[\frac{1}{r} \frac{\partial}{\partial r} (ru_r) + \frac{\partial u_z}{\partial z} \right] + \frac{1}{r} \frac{\partial}{\partial r} \left[r \left(\frac{\partial u_r}{\partial z} + \varepsilon^2 \frac{\partial u_z}{\partial r} \right) \right] = 0$$

Thin-Film Model

- Expand variables

$$h \sim h_0(r, t) + \varepsilon^2 h_1(r, t), \quad \phi_n \sim \phi_{n0}(r, z, t) + \varepsilon^2 \phi_{n1}(r, z, t), \quad \text{etc.}$$

- Simplified leading-order model

$$\frac{1}{r} \frac{\partial}{\partial r} (r u_{r0}) + \frac{\partial u_{z0}}{\partial z} = (1 + \Psi_m) \phi_{n0} g_{b0}$$

$$\frac{\partial \phi_{n0}}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} (r u_{r0} \phi_{n0}) + \frac{\partial}{\partial z} (u_{z0} \phi_{n0}) = \phi_{n0} g_{b0} - \Psi_d \phi_{n0}$$

$$\frac{\partial^2 g_{s0}}{\partial z^2} = 0$$

$$\frac{\partial^2 g_{b0}}{\partial z^2} = 0$$

$$\frac{\partial^2 u_{r0}}{\partial z^2} = 0$$

$$-\frac{\partial p_0}{\partial z} + \frac{1}{3} \frac{\partial}{\partial z} \left[\frac{1}{r} \frac{\partial}{\partial r} (r u_{r0}) + \frac{\partial u_{z0}}{\partial z} \right] + \frac{\partial^2 u_{z0}}{\partial z^2} = 0$$

Thin-Film Model

- Integrating across biofilm depth eliminates z dependence

$$\bar{\phi}_n = \frac{1}{h} \int_0^h \phi_n dz.$$

- Applying BCs gives a 1D system for $r \in [0, S(t)]$

$$\frac{\partial h_0}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} (r u_{r0} h_0) = (1 + \Psi_m) \bar{\phi}_{n0} g_{b0} h_0$$

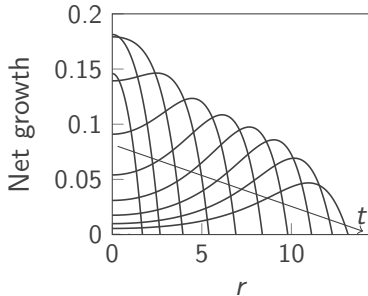
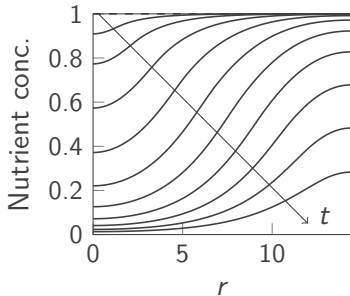
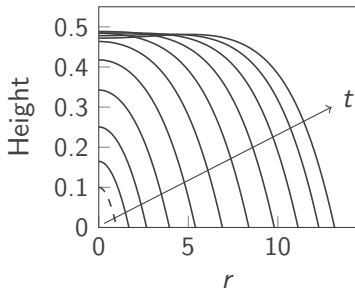
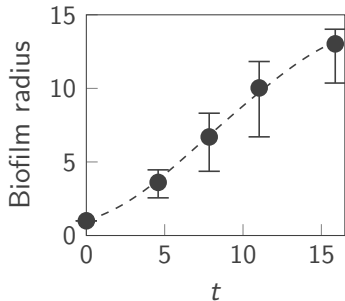
$$\frac{\partial \bar{\phi}_{n0}}{\partial t} + u_{r0} \frac{\partial \bar{\phi}_{n0}}{\partial r} = \bar{\phi}_{n0} [g_{b0} - \Psi_d - (1 + \Psi_m) \bar{\phi}_{n0} g_{b0}]$$

$$\frac{\partial g_{s0}}{\partial t} = D \left[\frac{1}{r} \frac{\partial}{\partial r} \left(r \frac{\partial g_{s0}}{\partial r} \right) - Q_s (g_{s0} - g_{b0}) \right]$$

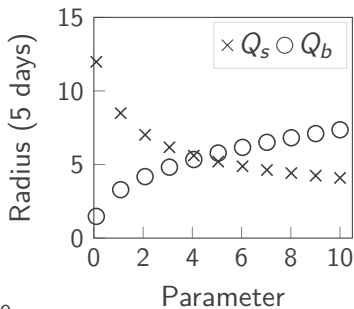
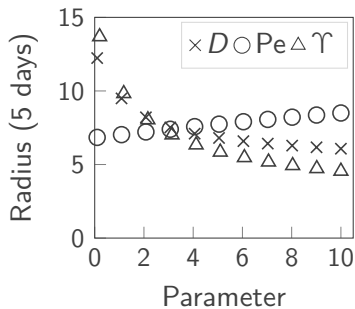
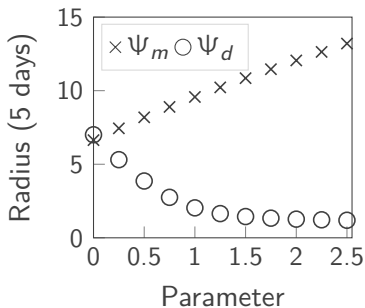
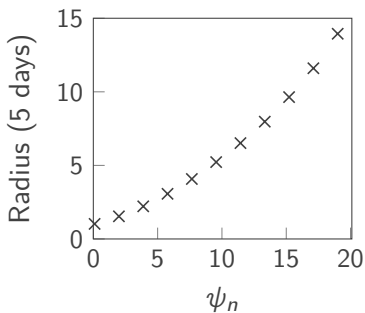
$$\text{Pe} \left[h_0 \frac{\partial g_{b0}}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} (r u_{r0} (1 - \bar{\phi}_{n0}) g_{b0} h_0) \right] = \frac{1}{r} \frac{\partial}{\partial r} \left(r h_0 \frac{\partial g_{b0}}{\partial r} \right) + Q_b (g_{s0} - g_{b0}) - \Upsilon \bar{\phi}_{n0} g_{b0} h_0$$

$$4 \frac{\partial}{\partial r} \left[\frac{h_0}{r} \frac{\partial}{\partial r} (r u_{r0}) \right] - 2 \frac{u_{r0}}{r} \frac{\partial h_0}{\partial r} = 2 (1 + \Psi_m) \frac{\partial}{\partial r} (\bar{\phi}_{n0} g_{b0} h_0) - \gamma^* h_0 \frac{\partial}{\partial r} \left[\frac{1}{r} \frac{\partial}{\partial r} \left(r \frac{\partial h_0}{\partial r} \right) \right]$$

Numerical Solutions



Effect of Parameters on Expansion Speed

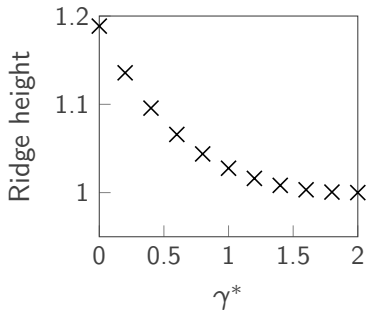
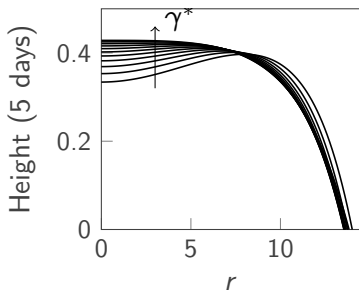


What About Surface Tension?

- In different experiments, yeast colonies can contain ridges⁶



- Surface tension does not affect biofilm size, but can inhibit ridge formation



⁶J. Maršíková et al., *BMC Genom.* 18 (2017), pp. 1–16.

Summary

- Yeast biofilms are a leading cause of bloodstream infections
- We modelled two hypothesised biofilm growth mechanisms
 - Nutrient-limited growth
 - Sliding motility
- Reaction–diffusion model with nonlinear degenerate cell diffusion could explain expansion speed and floral pattern⁷
- Two-phase thin-film fluid model for sliding motility predicts expansion in greater detail⁸
- Future work: 2D solutions to the fluid model

Acknowledgements:

- Supervisors and co-authors
- Funding: A. F. Pillow Trust and Research Training Program

⁷A. Tam et al., *J. Theor. Biol.* 448 (2018), pp. 122–141.

⁸A. Tam et al., *Proc. Royal Soc. A* 475 (2019), 20190175.

UQ Project: Actomyosin Networks

- Actin and myosin interactions in the cortex govern cell shape, movement, and division
- In experiments, disordered actomyosin networks contract
- Mechanisms of contractile stress generation currently disputed

