

A Thin-Film Extensional Flow Model for Biofilm Expansion by Sliding Motility

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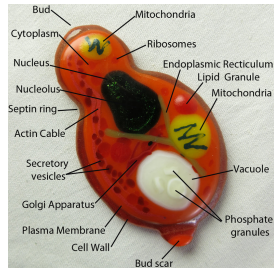
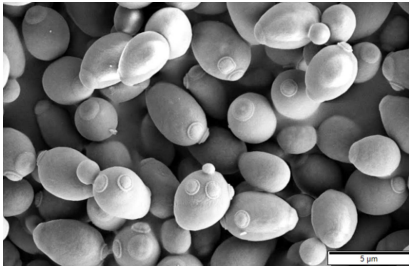
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Yeast

- Single-cell fungi ($\sim 4 \mu\text{m}$ diameter)
 - Food and drink
 - Waste management and biofuel production
- Baker's yeast (*S. cerevisiae*) is a common model organism
 - Similar to plant and animal cells
 - First eukaryotic genome to be sequenced
 - Helps develop anti-fungals and understand (cancer) cell division



Fungal Infections

- Pathogenic yeasts colonise medical devices and cause infections
 - Resist antimicrobial therapy — surgery often needed
 - Dangerous to immunocompromised people
 - Affects 1–2% of ICU patients, with up to 40% mortality rate¹
- Emerging pathogen *C. auris*: Japan 2009, 5 continents since
 - Highly resistant and difficult to diagnose

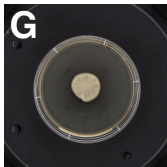
The image is a screenshot of a BBC News article. At the top, there is a navigation bar with the BBC logo, a 'BBC Account' link, a 'Menu' dropdown, and a search bar. Below this is a red banner with the word 'NEWS' in white. The article title is 'Candida auris: The new superbug on the block'. The author is 'By Lena Ciric' and the publication date is '17 August 2019'. To the right of the text is a world map titled 'Countries reporting cases of Candida auris, as of 31 May, 2019'. The map uses a legend where red indicates 'Multiple cases' and yellow indicates 'Single cases'. Red areas are visible in North America, Europe, and parts of Asia and Africa. Yellow areas are scattered in South America, Africa, and Asia. The source is cited as 'Source: CDC'.

- We seek common mechanisms underlying yeast biofilm growth

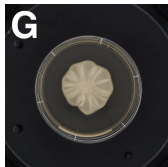
¹P. G. Pappas et al., *Nat. Rev. Dis. Primers* 4 (2018), 18026.

Yeast Biofilms

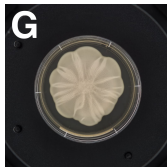
- Pathogenic yeasts form **biofilms**: sticky communities of cells and fluid existing on surfaces
 - Assist nutrient transport
 - Provide physical barrier to anti-fungals
- Lab-grown biofilms of baker's yeast form a floral pattern



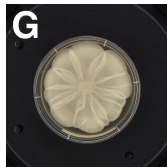
(a) Day 3



(b) Day 5



(c) Day 7

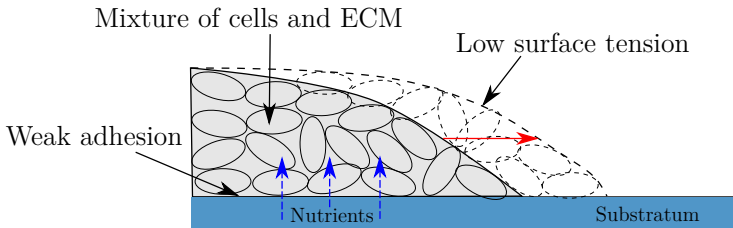


(d) Day 10

- Mechanisms of growth only understood qualitatively

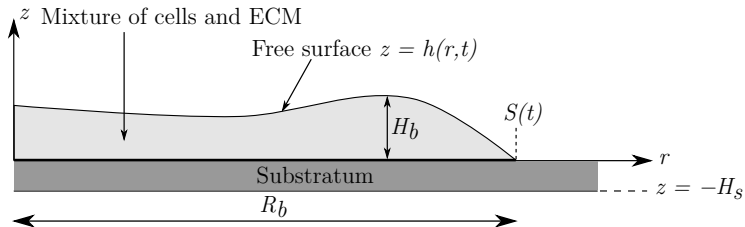
Sliding Motility

- **Hypothesis:** yeast biofilms expand by sliding motility²
 - Yeast adheres weakly to substratum — enables radial growth as cells proliferate
 - Biofilm takes up nutrients from the substratum
 - Nutrient consumption produces new cells and extracellular fluid
 - Cells and fluid spread passively as a unit



²T. B. Reynolds and G. R. Fink, *Science* 291 (2001), pp. 878–881.

Two-Phase Fluid Model



- Axisymmetric cylindrical geometry.
 - Biofilm occupies $0 \leq r \leq S(t)$ and $0 \leq z \leq h(r, t)$
- Biofilm is a mixture of two Newtonian viscous fluid phases:
 - Living cells $\phi_n(r, z, t)$ and ECM $\phi_m(r, z, t)$, with $\phi_n + \phi_m = 1$
 - Similar physical properties: $\rho_n = \rho_m$, $\mu_n = \mu_m$, etc.
 - Large interphase drag: $\mathbf{u}_n = \mathbf{u}_m$
- Thin aspect ratio

$$\frac{H_s}{R_b} = \varepsilon \ll 1, \quad \frac{H_b}{R_b} = \mathcal{O}(\varepsilon)$$

Governing Equations

- Mass balance (fluid phases)

$$\frac{\partial \phi_n}{\partial t} + \nabla \cdot (\phi_n \mathbf{u}) = \psi_n \phi_n g_b - \psi_d \phi_n$$

$$\frac{\partial \phi_m}{\partial t} + \nabla \cdot (\phi_m \mathbf{u}) = \psi_m \phi_n g_b + \psi_d \phi_n$$

- Mass balance (nutrients in the **substratum** and **biofilm**)

$$\frac{\partial g_s}{\partial t} = D_s \nabla^2 g_s$$

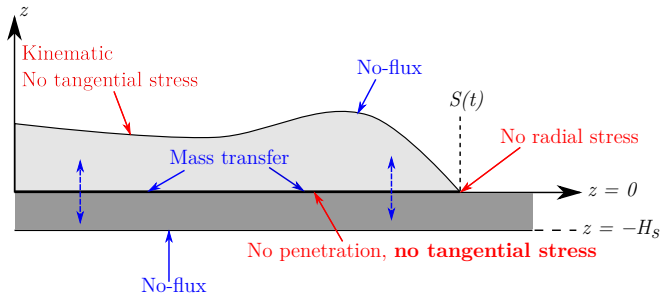
$$\frac{\partial g_b}{\partial t} + \nabla \cdot (g_b \phi_m \mathbf{u}) = D_b \nabla^2 g_b - \eta \phi_n g_b$$

- Momentum balance (fluid mixture), $\text{Re} \approx 0.001$

$$\nabla \cdot \boldsymbol{\sigma} = \mathbf{0}$$

Boundary Conditions

- Boundary conditions for **nutrients** and **fluids** close the model



- Nutrient transfer conditions on $z = 0$:

$$D_s \frac{\partial g_s}{\partial z} = -Q (g_s - g_b), \quad D_b \frac{\partial g_b}{\partial z} = -Q (g_s - g_b)$$

- No tangential stress** on substratum models weak adhesion
- Free surface normal stress proportional to local curvature:

$$\hat{\mathbf{n}} \cdot (\phi_\alpha \boldsymbol{\sigma} \cdot \hat{\mathbf{n}}) = -\gamma \kappa \quad \text{on} \quad z = h$$

Extensional Flow Scaling

- Scaling based on relevant physics
 - Thin biofilm (aspect ratio $\varepsilon \ll 1$)
 - Low surface tension
 - Nutrient-limited growth
- Variables

$$(r, z) = (R_b \hat{r}, \varepsilon R_b \hat{z}), \quad (u_r, u_z) = (\psi_n G R_b \hat{u}_r, \varepsilon \psi_n G R_b \hat{u}_z),$$

$$t = \frac{\hat{t}}{\psi_n G}, \quad g_s = G \hat{g}_s, \quad g_b = G \hat{g}_b, \quad p = \psi_n G \mu \hat{p}$$

- Parameters (estimated based on experiments)

$$\Psi_m = \frac{\psi_m}{\psi_n} = 0.11, \quad \Psi_d = \frac{\psi_d G}{\psi_n} = 0, \quad \gamma^* = \frac{\varepsilon \gamma}{\psi_n G R_b \mu} = 0,$$

$$D = \frac{D_s}{\psi_n G R_b^2} = 4.34, \quad \text{Pe} = \frac{\psi_n G R_b^2}{D_b} = 0.95, \quad \Upsilon = \frac{\eta R_b^2}{D_b} = 3.15,$$

$$Q_s = \frac{Q R_b}{\varepsilon D_s} = 2.09, \quad Q_b = \frac{Q R_b}{\varepsilon D_b} = 8.65$$

Thin-Film Model

- Expand variables

$$h \sim h_0(r, t) + \varepsilon^2 h_1(r, t), \quad \phi_n \sim \phi_{n0}(r, z, t) + \varepsilon^2 \phi_{n1}(r, z, t), \quad \text{etc.}$$

- Dimensionless model (dropping hats)

$$\frac{1}{r} \frac{\partial}{\partial r} (ru_r) + \frac{\partial u_z}{\partial z} = (1 + \Psi_m) \phi_n g_b$$

$$\frac{\partial \phi_n}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} (ru_r \phi_n) + \frac{\partial}{\partial z} (u_z \phi_n) = \phi_n g_b - \Psi_d \phi_n$$

$$\frac{\partial g_s}{\partial t} = D \left[\frac{1}{r} \frac{\partial}{\partial r} \left(r \frac{\partial g_s}{\partial r} \right) + \frac{1}{\varepsilon^2} \frac{\partial^2 g_s}{\partial z^2} \right]$$

$$\text{Pe} \left(\frac{\partial g_b}{\partial t} + \nabla \cdot [(1 - \phi_n) g_b \mathbf{u}] \right) = \frac{1}{r} \frac{\partial}{\partial r} \left(r \frac{\partial g_b}{\partial r} \right) + \frac{1}{\varepsilon^2} \frac{\partial^2 g_b}{\partial z^2} - \Upsilon \phi_n g_b$$

$$-\frac{\partial p}{\partial r} + \frac{2}{r} \frac{\partial}{\partial r} \left(r \frac{\partial u_r}{\partial r} \right) - \frac{2}{3} \frac{\partial}{\partial r} \left[\frac{1}{r} \frac{\partial}{\partial r} (ru_r) + \frac{\partial u_z}{\partial z} \right] + \frac{\partial}{\partial z} \left(\frac{\partial u_z}{\partial r} + \frac{1}{\varepsilon^2} \frac{\partial u_r}{\partial z} \right) - \frac{2}{r^2} u_r = 0$$

$$-\frac{\partial p}{\partial z} + 2 \frac{\partial^2 u_z}{\partial z^2} - \frac{2}{3} \frac{\partial}{\partial z} \left[\frac{1}{r} \frac{\partial}{\partial r} (ru_r) + \frac{\partial u_z}{\partial z} \right] + \frac{1}{r} \frac{\partial}{\partial r} \left[r \left(\frac{\partial u_r}{\partial z} + \varepsilon^2 \frac{\partial u_z}{\partial r} \right) \right] = 0$$

Thin-Film Model

- Expand variables

$$h \sim h_0(r, t) + \varepsilon^2 h_1(r, t), \quad \phi_n \sim \phi_{n0}(r, z, t) + \varepsilon^2 \phi_{n1}(r, z, t), \quad \text{etc.}$$

- Simplified leading-order model

$$\frac{1}{r} \frac{\partial}{\partial r} (r u_{r0}) + \frac{\partial u_{z0}}{\partial z} = (1 + \Psi_m) \phi_{n0} g_{b0}$$

$$\frac{\partial \phi_{n0}}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} (r u_{r0} \phi_{n0}) + \frac{\partial}{\partial z} (u_{z0} \phi_{n0}) = \phi_{n0} g_{b0} - \Psi_d \phi_{n0}$$

$$\frac{\partial^2 g_{s0}}{\partial z^2} = 0$$

$$\frac{\partial^2 g_{b0}}{\partial z^2} = 0$$

$$\frac{\partial^2 u_{r0}}{\partial z^2} = 0$$

$$-\frac{\partial p_0}{\partial z} + \frac{1}{3} \frac{\partial}{\partial z} \left[\frac{1}{r} \frac{\partial}{\partial r} (r u_{r0}) + \frac{\partial u_{z0}}{\partial z} \right] + \frac{\partial^2 u_{z0}}{\partial z^2} = 0$$

Thin-Film Model

- Integrating across biofilm depth eliminates z dependence

$$\bar{\phi}_n = \frac{1}{h} \int_0^h \phi_n dz.$$

- Applying BCs gives a 1D system for $r \in [0, S(t)]$

$$\frac{\partial h_0}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} (r u_{r0} h_0) = (1 + \Psi_m) \bar{\phi}_{n0} g_{b0} h_0$$

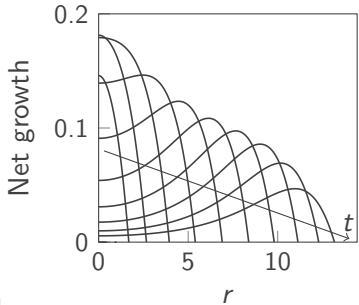
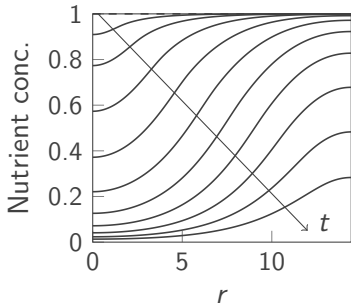
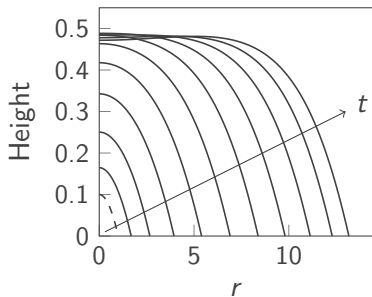
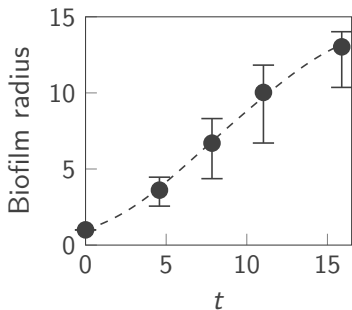
$$\frac{\partial \bar{\phi}_{n0}}{\partial t} + u_{r0} \frac{\partial \bar{\phi}_{n0}}{\partial r} = \bar{\phi}_{n0} [g_{b0} - \Psi_d - (1 + \Psi_m) \bar{\phi}_{n0} g_{b0}]$$

$$\frac{\partial g_{s0}}{\partial t} = D \left[\frac{1}{r} \frac{\partial}{\partial r} \left(r \frac{\partial g_{s0}}{\partial r} \right) - Q_s (g_{s0} - g_{b0}) \right]$$

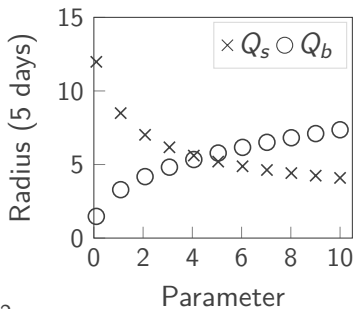
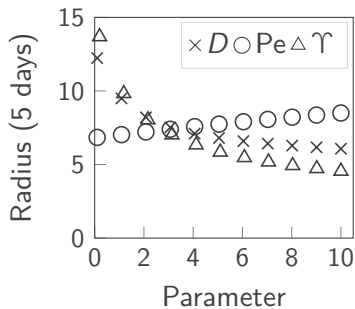
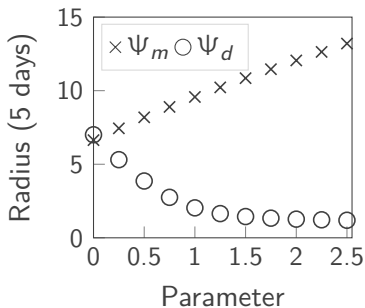
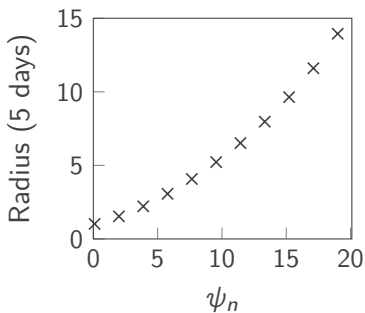
$$\text{Pe} \left[h_0 \frac{\partial g_{b0}}{\partial t} + \frac{1}{r} \frac{\partial}{\partial r} (r u_{r0} (1 - \bar{\phi}_{n0}) g_{b0} h_0) \right] = \frac{1}{r} \frac{\partial}{\partial r} \left(r h_0 \frac{\partial g_{b0}}{\partial r} \right) + Q_b (g_{s0} - g_{b0}) - \Upsilon \bar{\phi}_{n0} g_{b0} h_0$$

$$4 \frac{\partial}{\partial r} \left[\frac{h_0}{r} \frac{\partial}{\partial r} (r u_{r0}) \right] - 2 \frac{u_{r0}}{r} \frac{\partial h_0}{\partial r} = 2 (1 + \Psi_m) \frac{\partial}{\partial r} (\bar{\phi}_{n0} g_{b0} h_0) - \gamma^* h_0 \frac{\partial}{\partial r} \left[\frac{1}{r} \frac{\partial}{\partial r} \left(r \frac{\partial h_0}{\partial r} \right) \right]$$

Numerical Solutions



Effect of Parameters on Expansion Speed

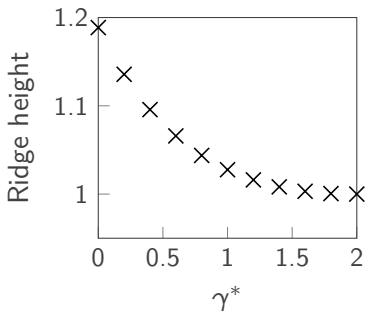
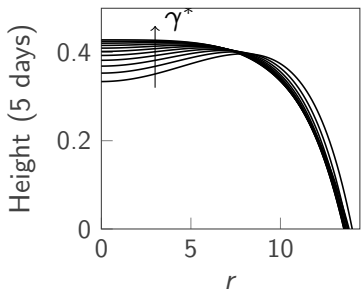


What About Surface Tension?

- In different experiments, yeast colonies can contain ridges³



- Surface tension represents cell-cell adhesion strength⁴
- Non-zero γ^* does not affect size, but inhibits ridge formation



³J. Maříková et al., *BMC Genom.* 18 (2017), pp. 1–16.

⁴G. Forgacs et al., *Biophys. J.* 74 (1998), pp. 2227–2234.

Summary

- Yeast biofilms are a leading cause of bloodstream infections
- Two-phase thin-film extensional flow model describes expansion by sliding motility⁵
- Model shows how to enhance/inhibit growth and ridge formation
- Future work: strong adhesion model, pattern formation

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- ANZIAM, Organisers, MBSIG
- Supervisors and Co-authors



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 @xelamaths

⁵A. Tam et al., *Proc. Royal Soc. A* 475 (2019), 20190175.